

Chapter 6



How cells grow



Introduction

Cell growth is the primary response of viable cells to substrates and nutrients.

Substrates/nutrients + cells \rightarrow products + more cells

$$\text{specific growth rate (h}^{-1}\text{), } \mu \equiv \frac{1}{X} \frac{dX}{dt}$$

X = cell mass concentration (g / L)

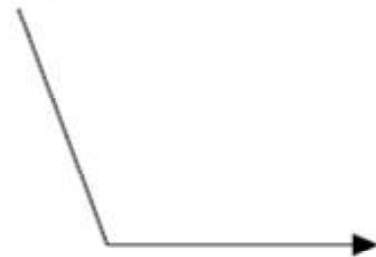
t = time (h)

Product formation is a secondary response.

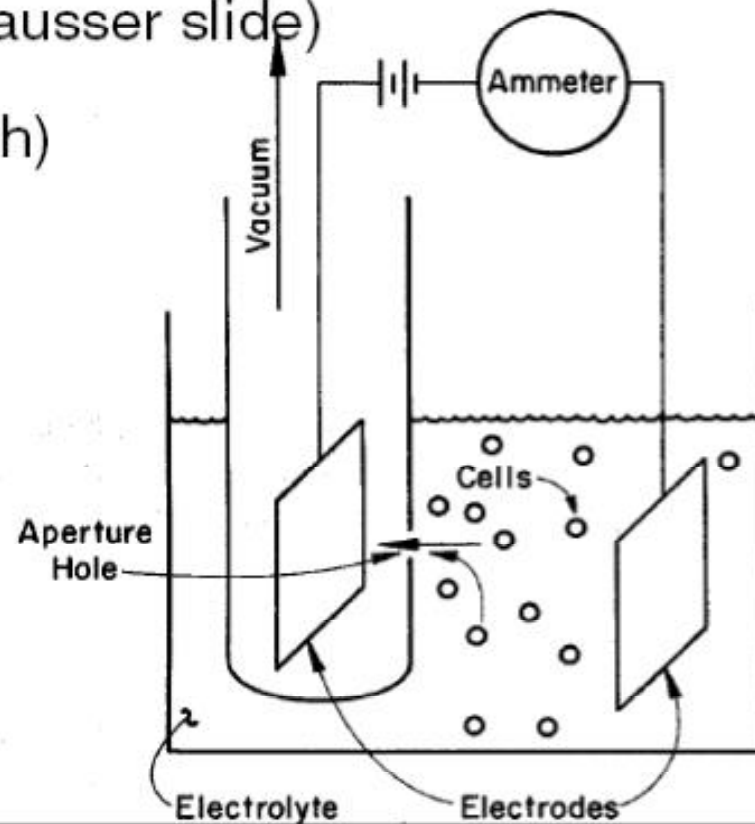
Determining cell concentration

1. Cell number concentration (cells/mL)

- a) hemocytometer (Petroff-Hausser slide)
- b) viable cell counts (petri dish)
- c) electronic particle counter

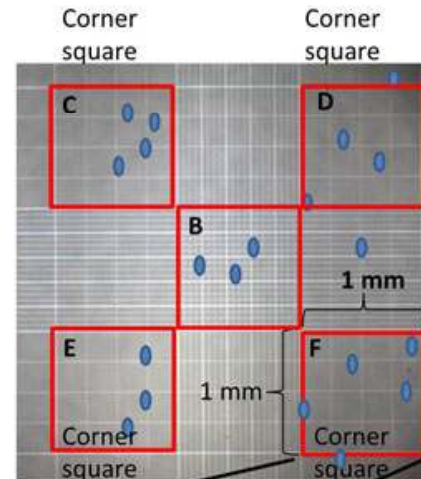
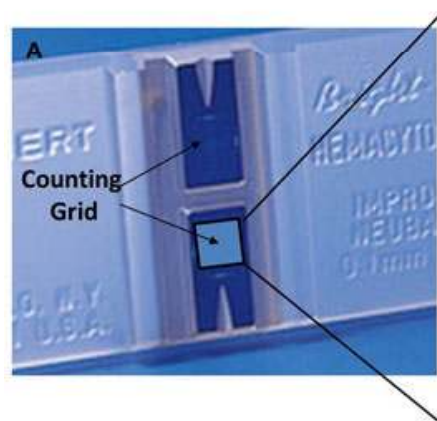
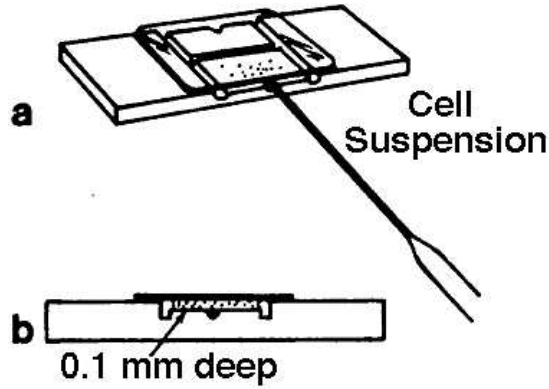


*"Bioprocess Engineering: Basic Concepts
Shuler and Kargi, Prentice Hall, 2002*



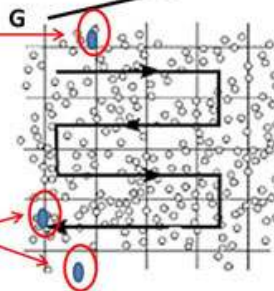
Determining cell concentration

Hemocytometer



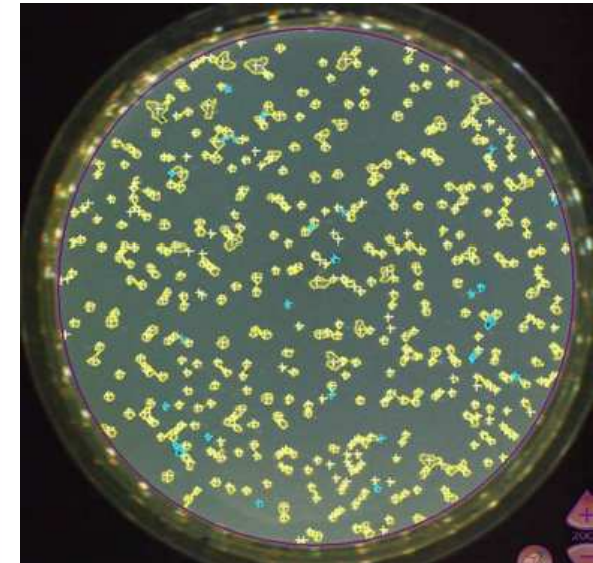
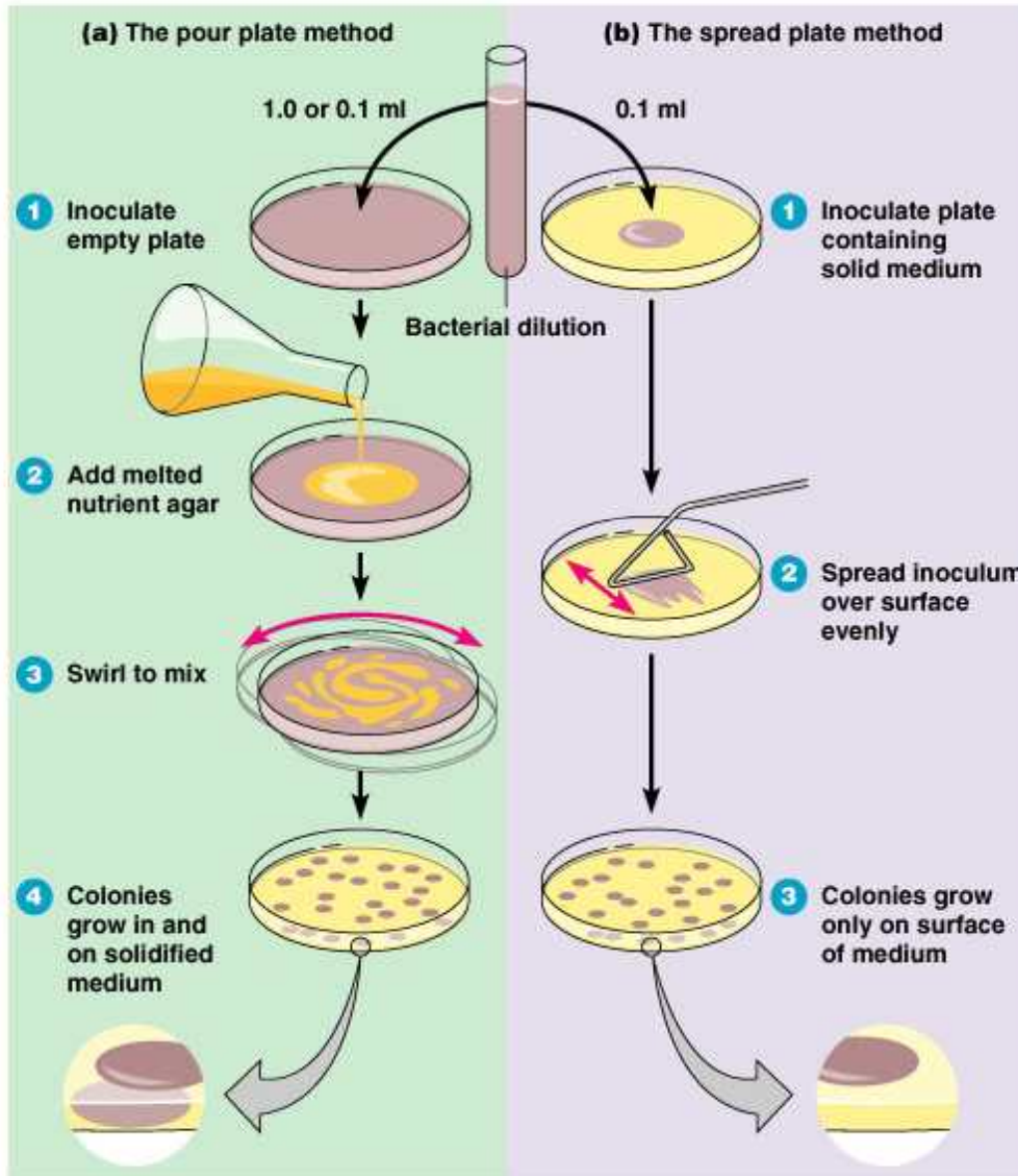
Cell touching the top ruling = in

Cell touching the left or bottom ruling = out



Determining cell concentration

Bacteria





Determining cell concentration (cont.)

2. Cell mass concentration (g/mL)

a) direct methods

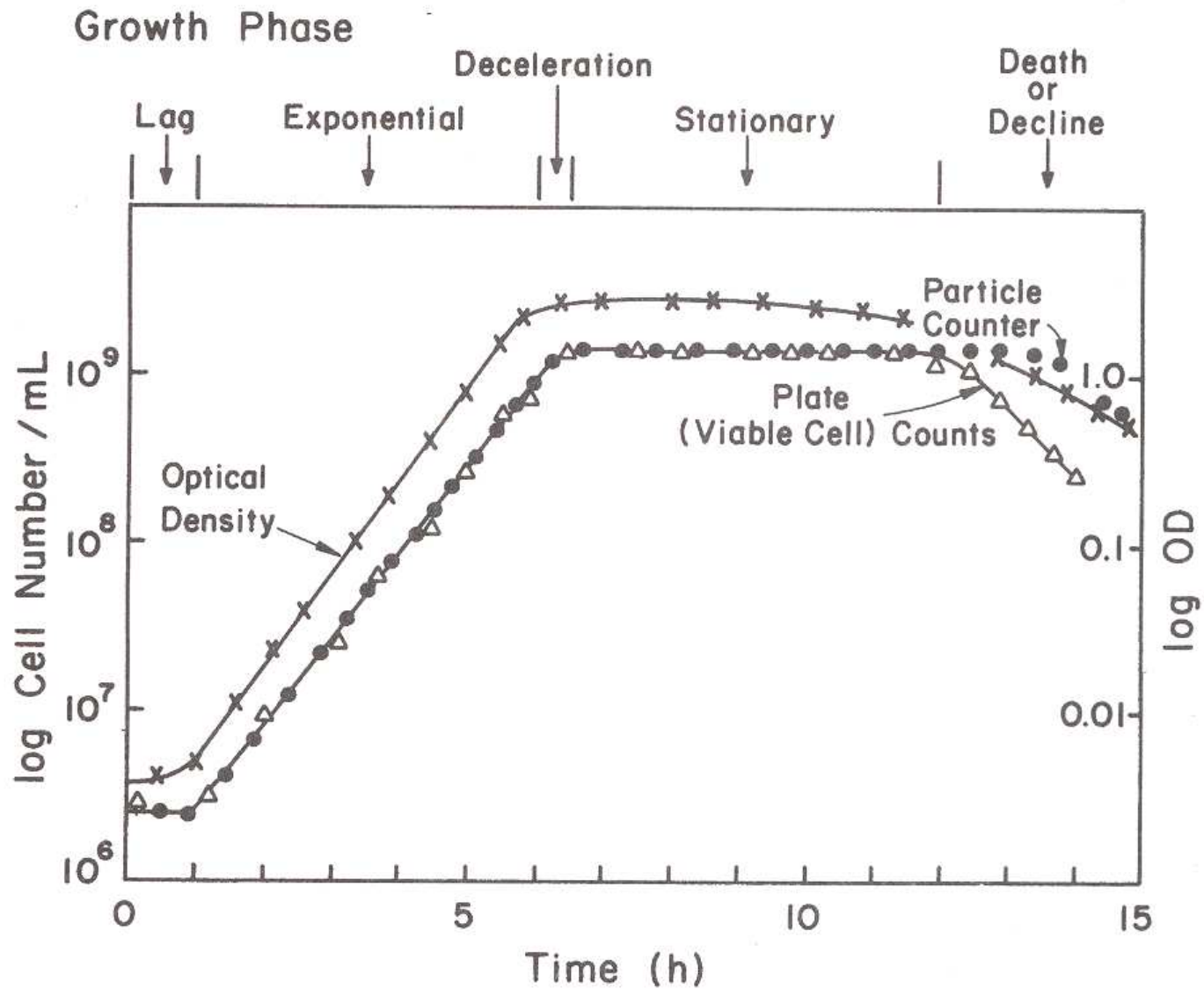
- dry weight (filtration or centrifugation)
- packed cell volume (centrifugation)
- optical density (light scattering, 600-700 nm)

b) indirect methods

→ measure biomolecule concentration and correlate to dry cell mass concentration.

(DNA, protein, ATP, NADH, product formation)

Cell growth kinetics in batch culture





Lag phase

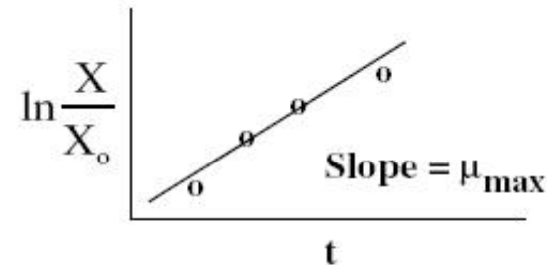
- no increase in cell numbers
- induction of enzymes to utilize | substrate(s)
- very important to decrease lag period to ↑ productivity
 - i. Inoculate with exponential phase cells
 - ii. Pre-acclimate inoculum in growth media
 - iii. Use high cell inoculum size

Exponential growth phase

1. Nutrient and substrate concentrations are high
2. Growth rate is independent of nutrient and substrate conc.
3. Cell number and mass concentrations increase exponentially

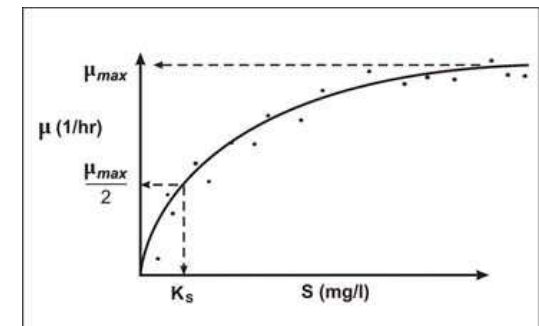
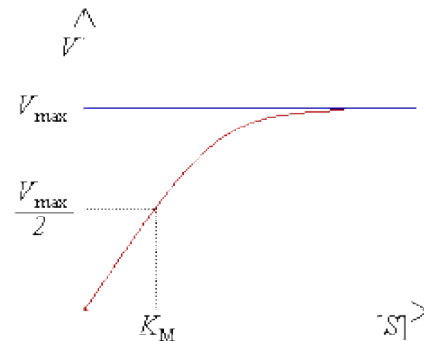
$$\frac{dX}{dt} = \mu_{\max} X, \quad X = X_0 \text{ at } t = 0$$

$$X = X_0 e^{\mu_{\max} t} \text{ or } \ln \frac{X}{X_0} = \mu_{\max} t$$



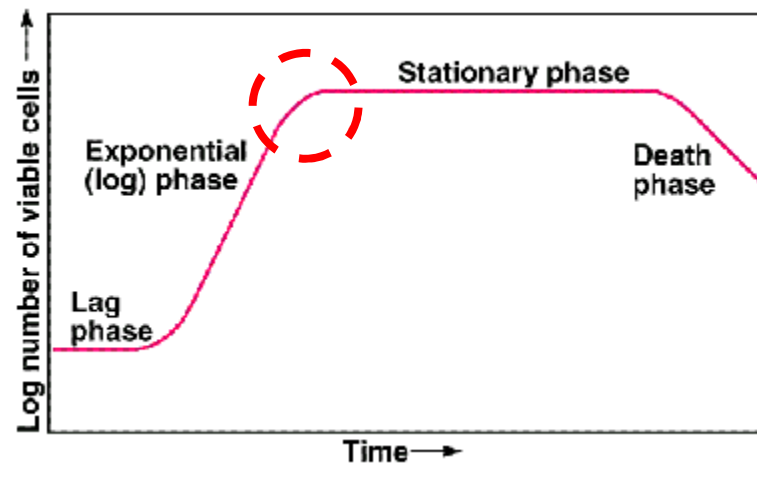
doubling time of cells (t_d), $\frac{X}{X_0} = 2 \Rightarrow \ln(2) = \mu_{\max} t_d$

$$t_d = \frac{\ln 2}{\mu_{\max}} \quad \text{or} \quad \mu_{\max} = \frac{\ln 2}{t_d}$$



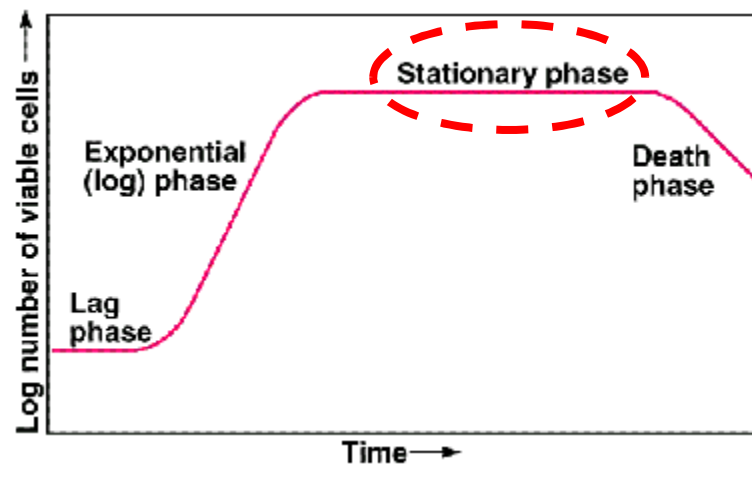
Deceleration phase

- depletion of one or more nutrients
- accumulation of toxic byproducts of growth



Stationary phase

- no net growth of cell numbers or cell mass (no cell division)
- cell growth rate = cell death rate
- *secondary metabolites* (products) produced
- removal of inhibitory compounds will result in further growth if additional substrate is provided



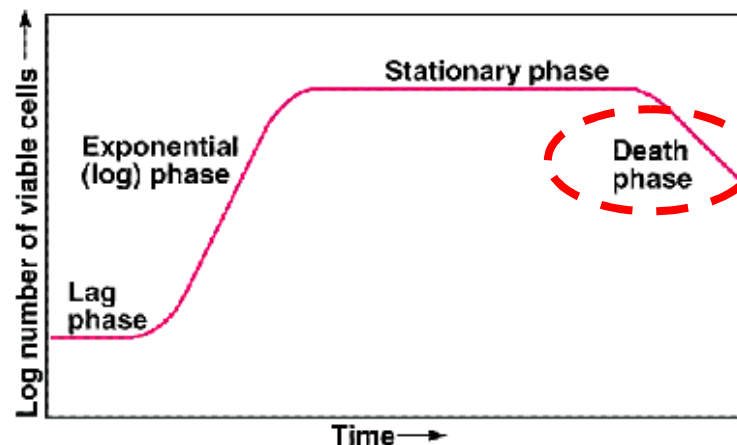
Death phase

1. Cell lysis (spillage) may occur
2. Rate of cell decline is first-order

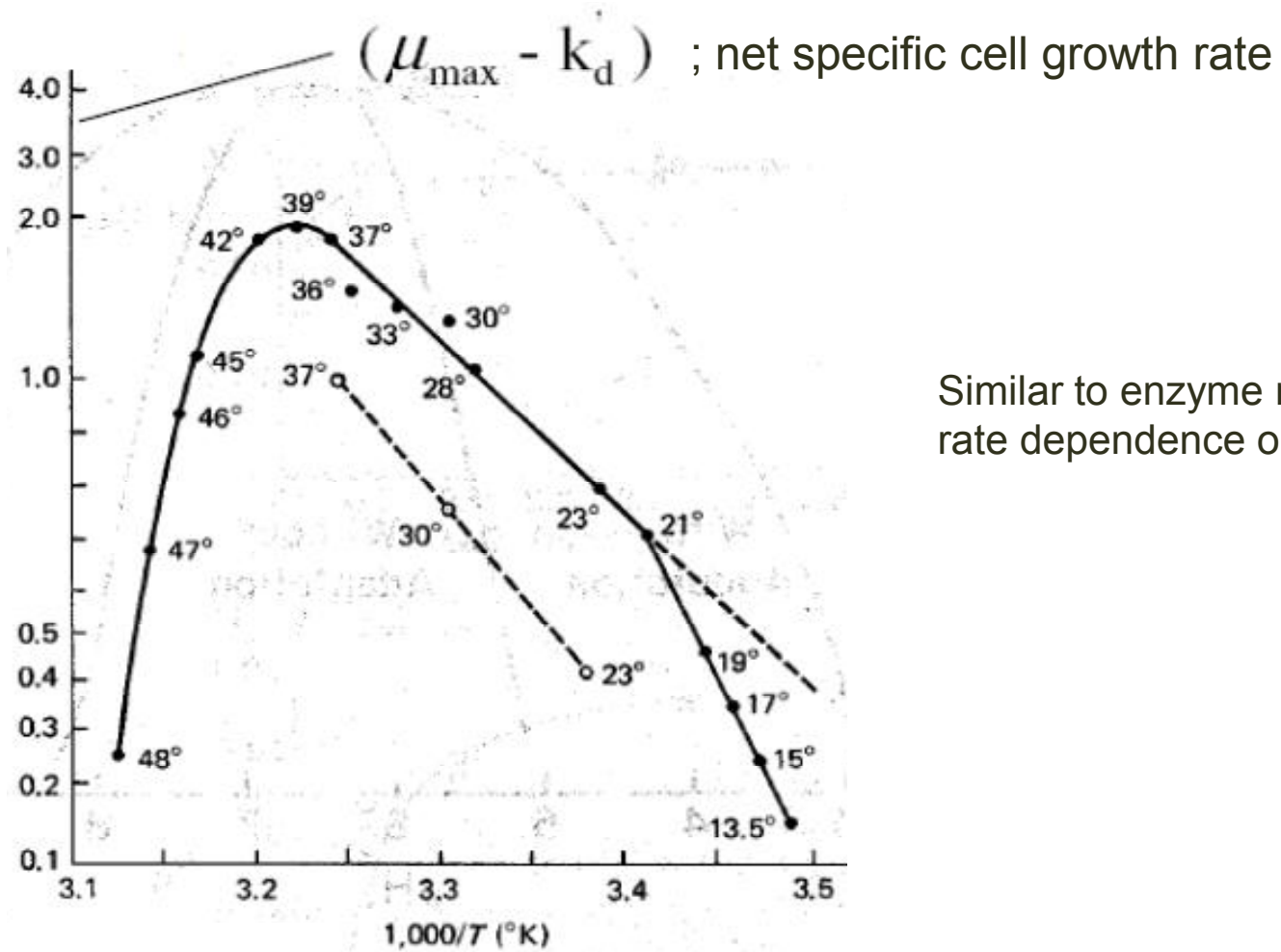
$$\frac{dX}{dt} = -k_d' X, \Rightarrow X = X_s \text{ at } t = 0 \quad k_d'; \text{ specific cell death rate}$$

$$X = X_s e^{-k_d' t} \quad \text{or} \quad \ln \frac{X}{X_s} = -k_d' t$$

3. Growth can be re-established by transferring to fresh media



Effects of temperature on cell growth



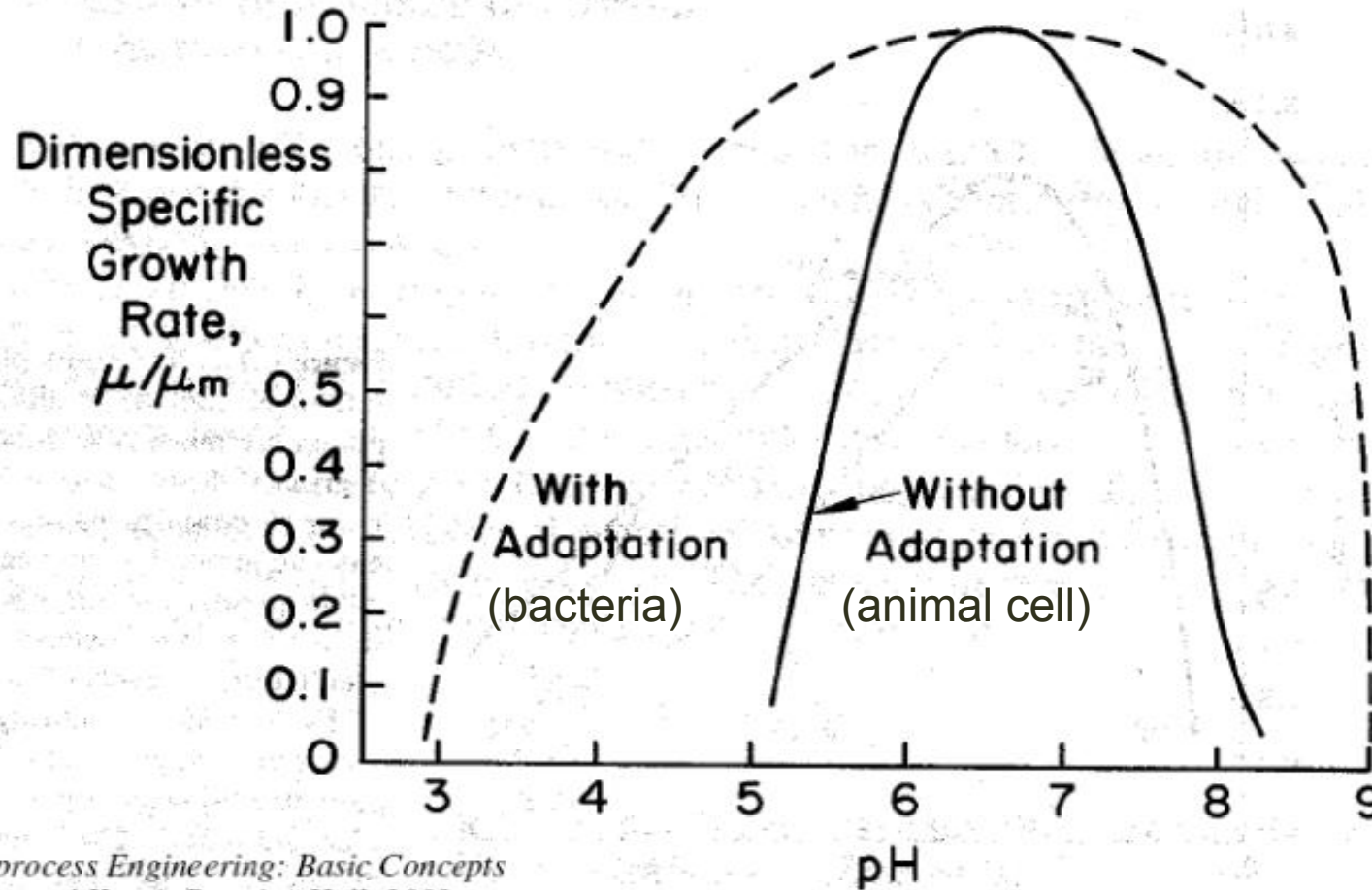
Similar to enzyme reaction rate dependence on T



pH effects

- acceptable pH is ± 1 to 2 pH units
- pH range varies by organism
 - bacteria (most) pH = 3 to 8
 - yeast pH = 3 to 6
 - plants pH = 5 to 6
 - animals pH = 6.5 to 7.5
- microorganism have the ability to control pH inside the cell, but this requires maintenance energy
- pH can change due to
 - utilization of substrates; NH_4^+ releases H^+ , NO_3^- consumes H^+
 - production of organic acids, amino acids, CO_2 , bases

pH effects (cont.)



Bioprocess Engineering: Basic Concepts
Shuler and Kargi, Prentice Hall, 2002

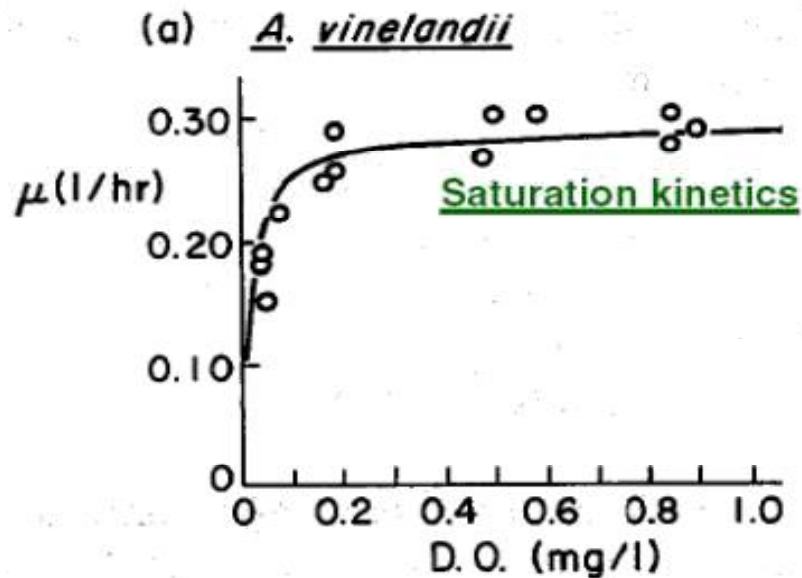


Effects of dissolved O₂ (DO)

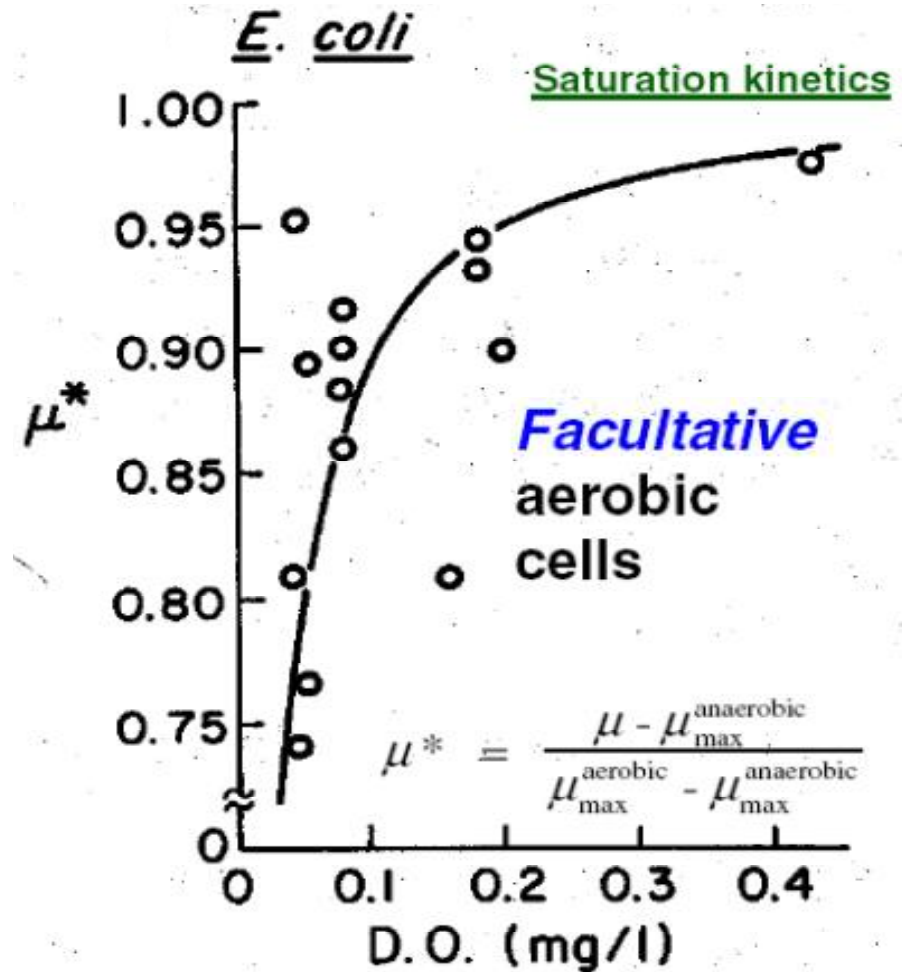
- O₂ may be a limiting substrate for aerobic fermentation, since O₂ is sparingly soluble in water
- critical O₂ concentration
5 to 10% of saturation ($\approx 7 \text{ mg O}_2/\text{L}$) for bacteria/yeast
- growth exhibits saturation kinetics with respect to O₂ concentration (see next page)

Effects of dissolved O₂ (cont.)

Obligate aerobic cells

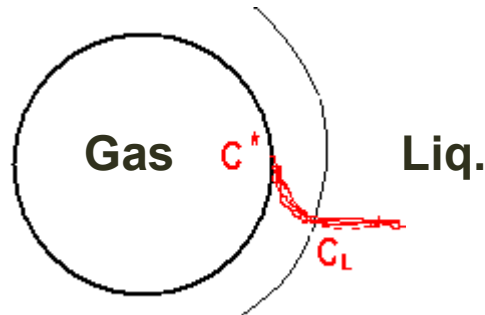


"Bioprocess Engineering: Basic Concepts
Shuler and Kargi, Prentice Hall, 2002



Oxygen transfer rate (OTR)

■ Oxygen transfer from gas to liquid



■ $OTR = k_L a (C^* - C_L)$

- OTR [mg O₂ / L / h]
- k_L : oxygen transfer coefficient (cm/h)
- a : gas-liquid interfacial area per unit vol. (cm²/cm³)
- $k_L a$: volumetric oxygen transfer coefficient (1/h)
- C^* : saturated DO concentration (mg/L)
- C_L : DO concentration in the broth (mg/L)

Oxygen uptake rate (OUR)

■ OUR from liquid to cell

$$\text{OUR} = q_{\text{O}_2} X = (\mu X) / Y_{\text{X/O}_2}$$

- OUR [mg O₂ / L / h]
- q_{O₂} : specific rate of oxygen consumption (mg O₂/g cell/h)
- Y_{X/O₂} : oxygen yield coefficient (g cell/g O₂)

■ When oxygen transfer is the rate-limiting step,

$$\text{OTR} (\rightarrow) = \text{OUR} (\rightarrow)$$

$$k_L a (C^* - C_L) = (\mu X) / Y_{\text{X/O}_2}$$

$$Y_{\text{X/O}_2} k_L a (C^* - C_L) = dX / dt$$

$$Y_{\text{X/O}_2} \cdot \text{OTR} = \text{cell growth rate}$$





Other effects on cell growth

→ dissolved CO₂ (DCO₂); too high or low DCO₂ is toxic

→ ionic strength (I); too high dissolved salts is inhibitory to membrane function (membrane transport of nutrients, osmotic pressure)

$$I = 1/2 \sum C_i Z_i^2$$

C_i = molar concentration of ion i

Z_i = ion charge

→ maximum non-inhibitory concentrations of substrates, products
glucose (100 g/L), ethanol (10 g/L), NH₄⁺ (5 g/L), ..