



# 10. Separation in Solution







# 1. Gel Filtration





# Gel Filtration

## ■ Other names

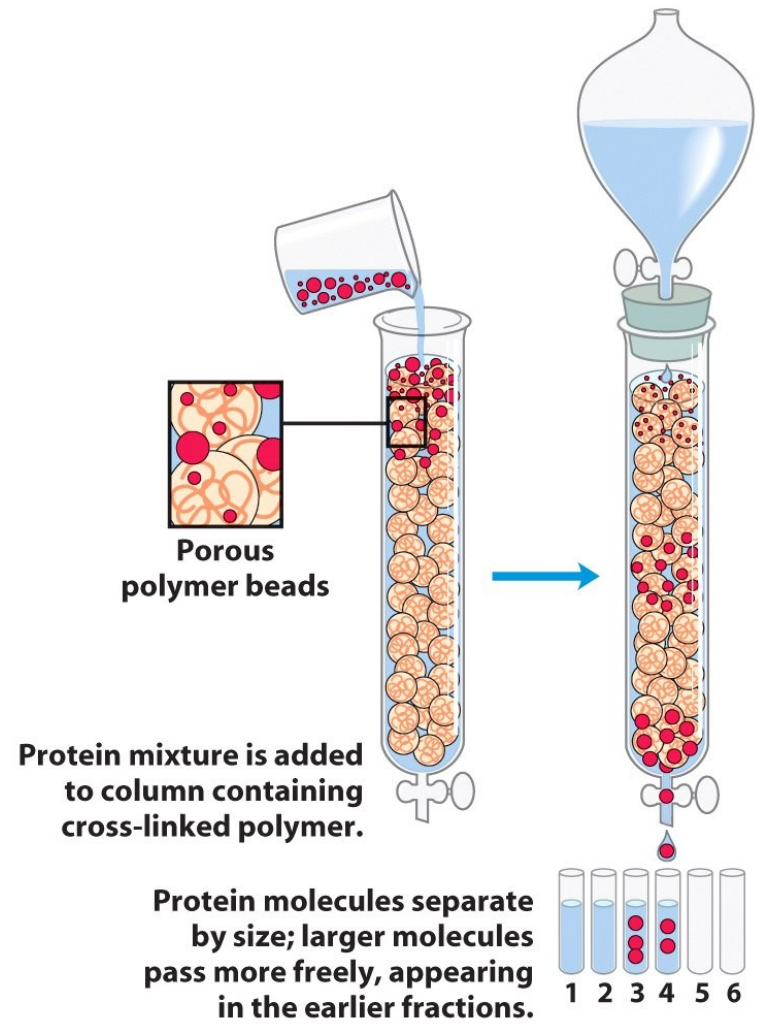
- Gel permeation
- Gel exclusion
- Molecular sieving

## ■ Principle

- Separation of protein according to differences in size
- Limited accessibility of protein to cross-linked porous matrix
  - Small molecules diffuse in and out of the pores → stay longer on the column
- No binding of protein to the matrix
  - Buffer condition does not affect resolution

## ■ Application

- Purification
- Molecular weight determination





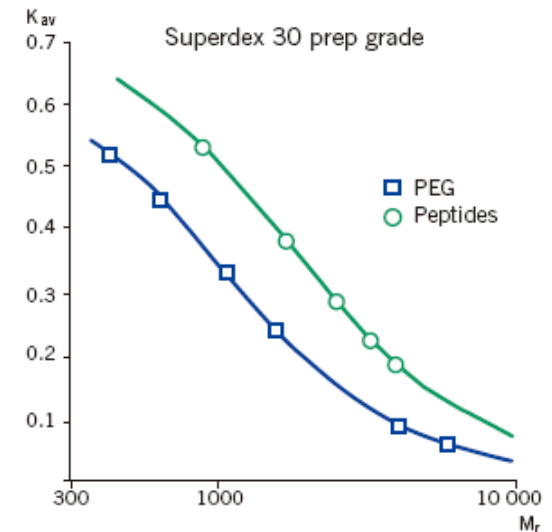
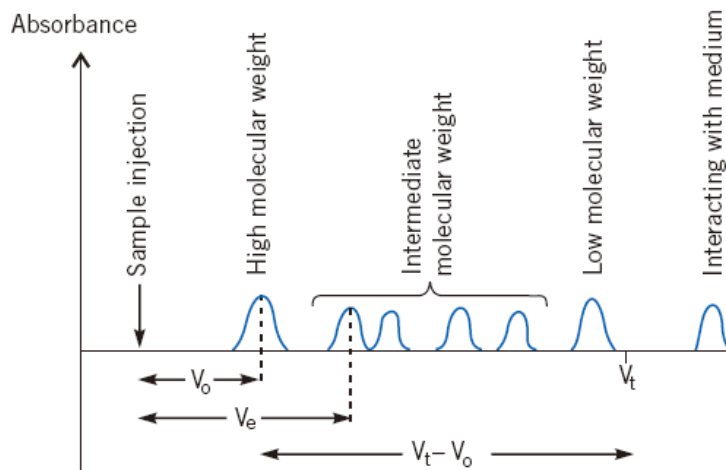
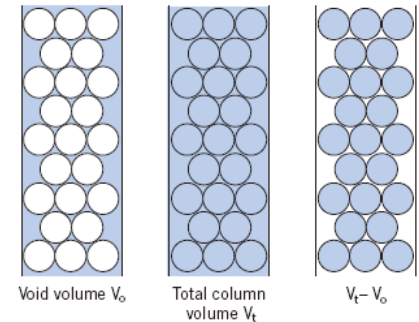
# Principles of Gel Filtration

## ■ Partition coefficient $K_{av}$

- Proportion of pores occupied by the molecule
- $K_{av} = (V_e - V_o) / (V_t - V_o)$
- Depending on protein behavior and the nature of the gel beads
- Independent of column size

## ■ $K_{av}$ for gel filtration chromatography

- 0 (High molecular weight protein)  $< K_{av} < 1$  (Low molecular weight protein)
- $K_{av} > 1$  : non-specific binding of the molecule
- $K_{av} < 0$  : Channeling in the chromatography bed → need repacking

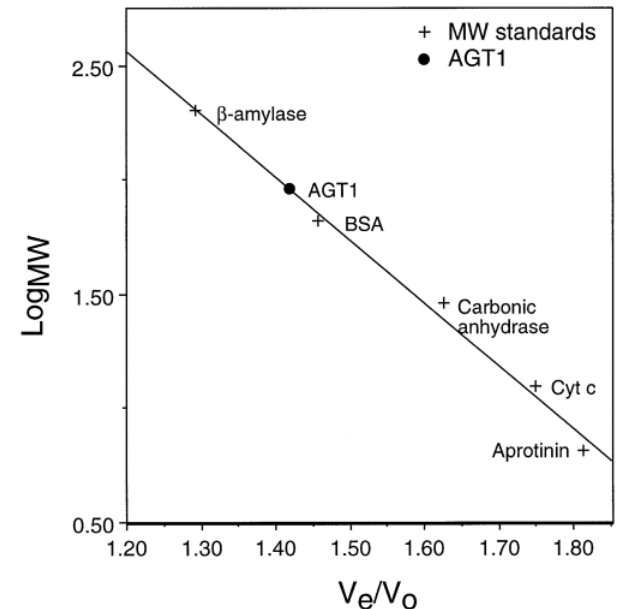




# Applications

## ■ Determination of protein molecular weight

- Using calibrations of elution volumes of known proteins
- Elution volume is related to stokes radius
  - A sphere with a hydrodynamic behavior equivalent to that of a particular irregular shaped particle
- For proteins with similar shape, calibration curve between  $\log MW$  and  $K_{av}$  can be used for to determine molecular weight of an unknown protein



## ■ Purification of proteins

- High resolution than the exact position of elution is important
- High resolution: 1/3~2/3 of separation range

$\beta$ -amylase: 200 kDa

BSA: 66 kDa

Carbonic anhydrase : 29 kDa

Cytochrome c :12.4 kDa

Aprotinin: 6.5 kDa

Liepman A. H., et al., 2001, The Plant Journal



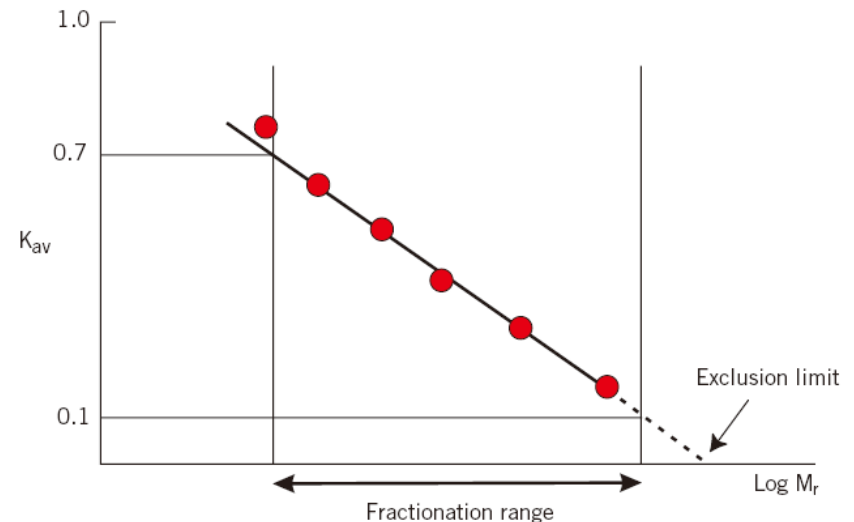
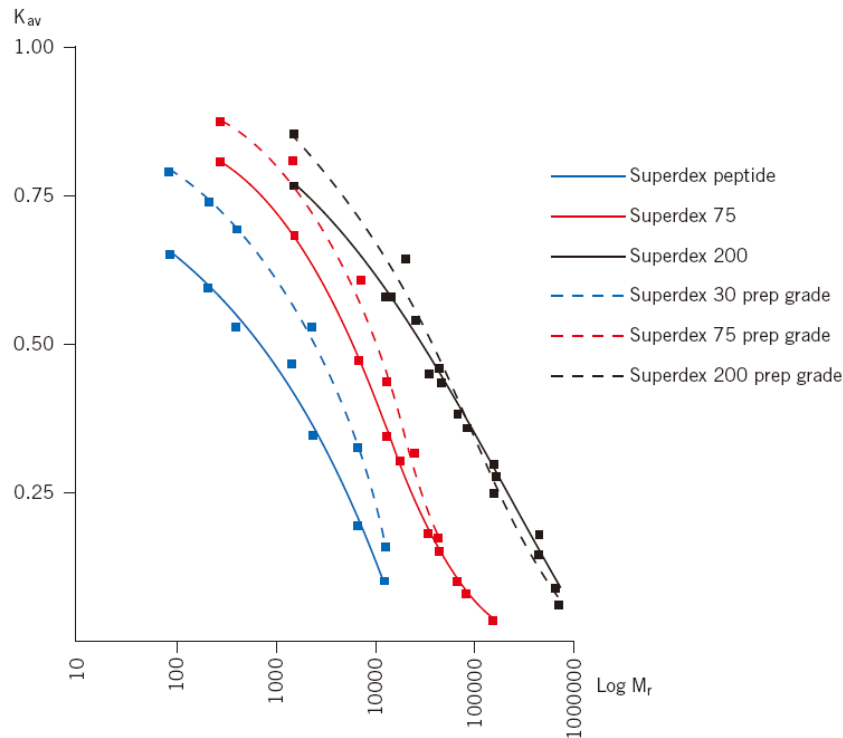
# Selection of Media

## ■ Depending on purpose

- High resolution fractionation
- Group separation

## ■ Depending on the size of the target protein

- The target protein should be within the fractionation range ( $K_{av}$  0.1 to 0.7)
- $M_w$  100 ~  $8 \times 10^8$





# Gel Filtration Media

## ■ Cross-linked dextran

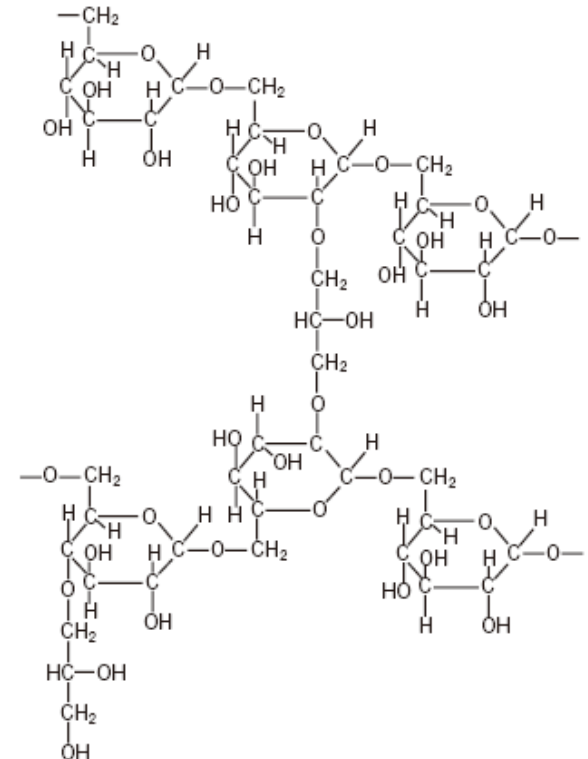
### ■ Sephadex

- Controlling the degree of cross-linking and particle size
- For rapid group separation of high and low molecular weight substances
  - Desalting, buffer exchange and sample clean up
  - e.g Sephadex G-25
    - » Excellent for removing salt and contaminant from protein of > 5 kDa

## ■ Cross-linked agarose : Higher porosity and strength

### ■ Superose

- Broad fractionation range, not suitable for industrial scale





# Gel Filtration Media

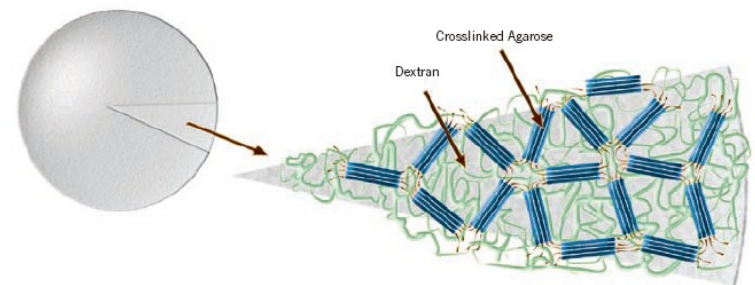
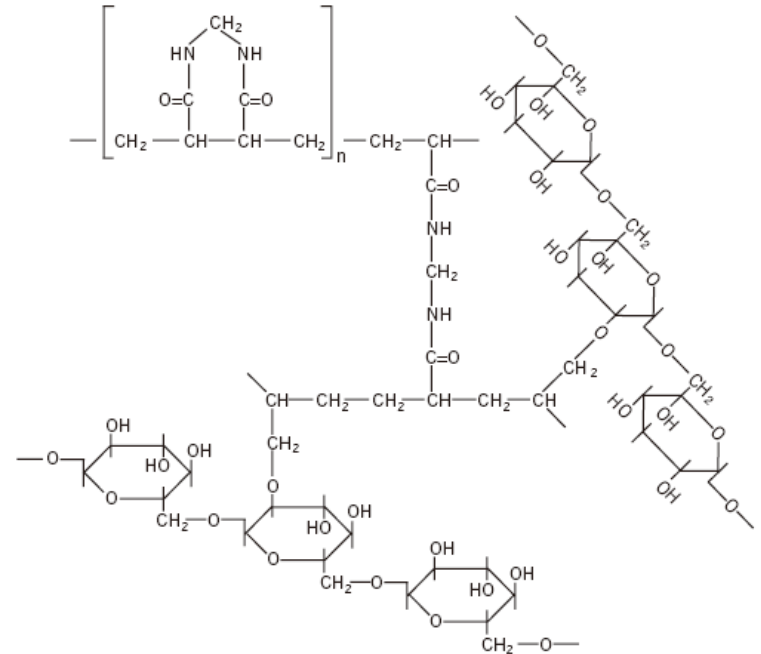
## ■ Composite

### ■ Sephacryl

- Cross-linked allyl dextran with N,N'-methylene bisacrylamide
- Fast, high recovery at laboratory and industrial scale

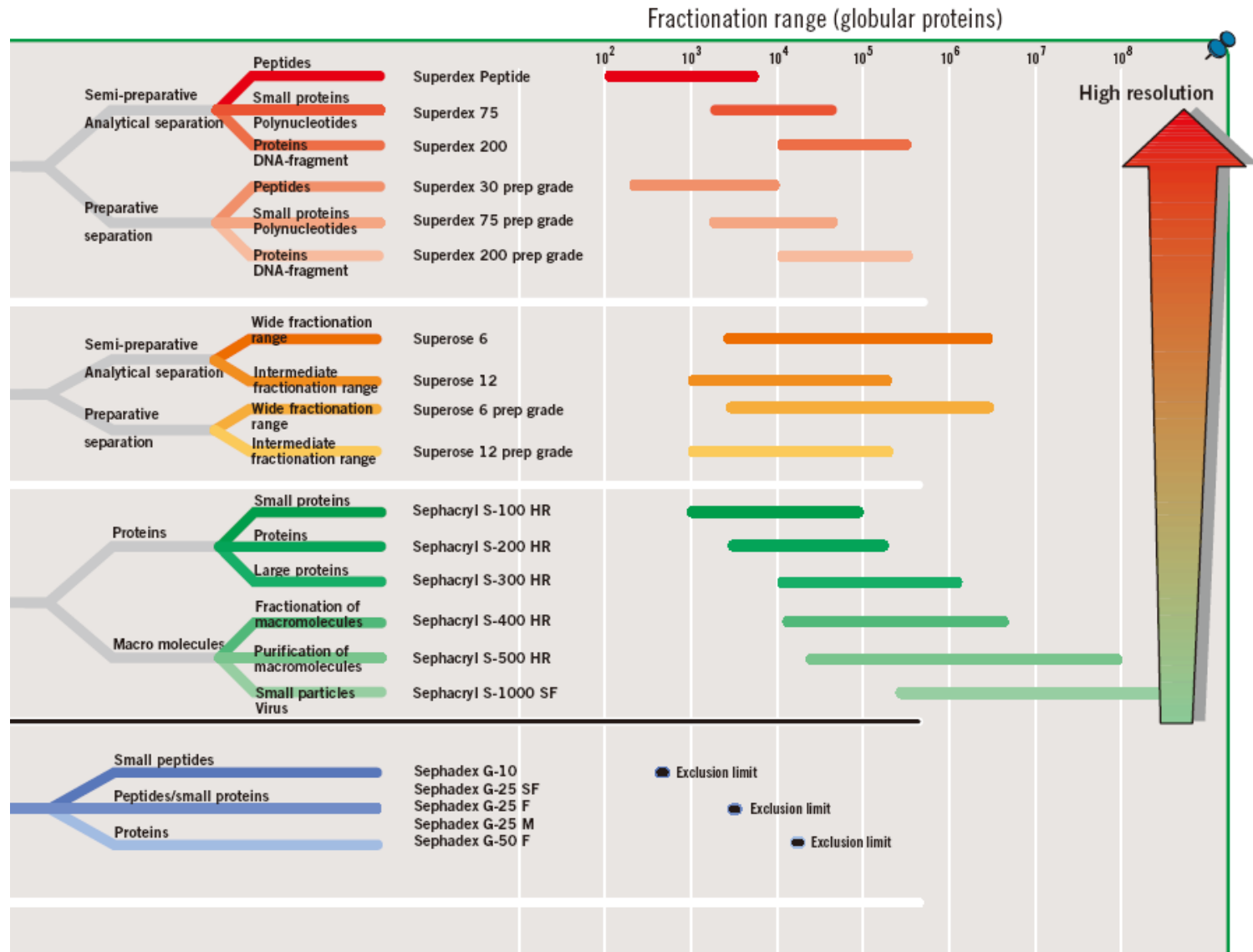
### ■ Superdex

- Highly cross-linked agarose + covalent attachment of dextran chain
- Very high stability
- Fast, high resolution and recovery





# Selection of Media





# Factors Affecting Resolution

## ■ Sample

- The ratio of sample volume to column volume
- Viscosity of the mobile phase

## ■ Flow rate

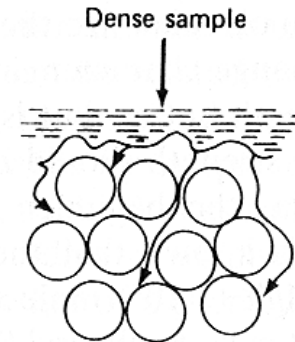
## ■ Column

- Column dimension or shape
  - Squat column : poor resolution by minor deviation of flow
  - Length ~ 20-40x of diameter
- Particle size
  - The smaller the better, but need higher pressure
- Particle size distribution
- Packing density
- Porosity of the particle

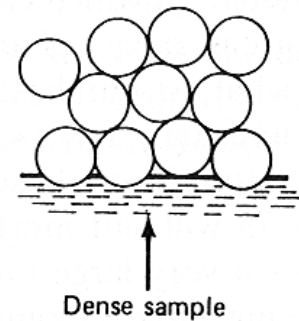
## ■ Gravitational instability

- Dense solution moves faster than expected

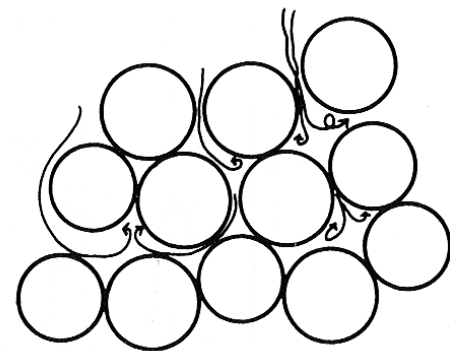
## ■ Turbulent flow between particles



(a)



(b)





# Factors Affecting Elution Pattern

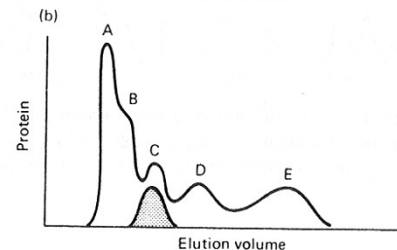
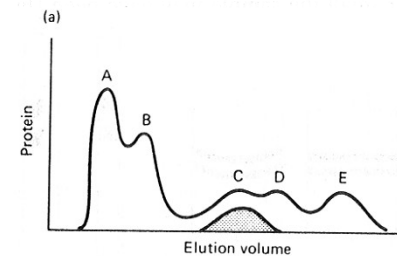
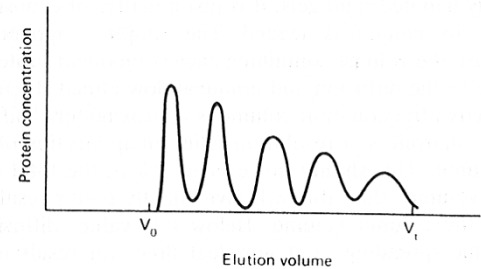
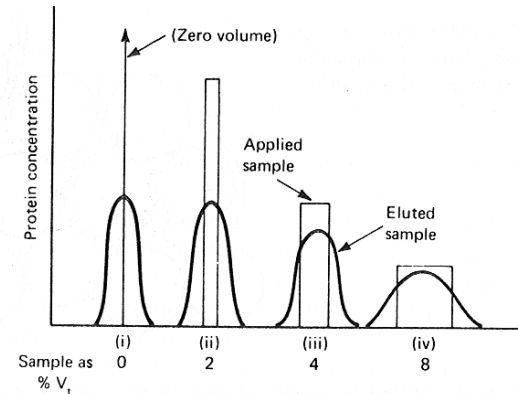
## ■ Sample volume

- 1~3% of column volume
- Smaller volume
  - Similar level of broadening because diffusion, natural chromatographic spreading, and nonideal flow
- Higher volume
  - Band broadening

## ■ Protein size

- Smaller proteins
  - High diffusion coefficient and longer stay in the column
  - More diffusion and band broadening

## ■ Pore size



Large pores

Small pores





## 2. Electrophoretic Methods





# Electrophoresis for Protein Purification

## ■ Electrophoresis

- Electrophoresis under constant pH current
- Isoelectric focusing
  - Separation in pH gradient
- Isotachopheresis
  - Variation of electric field according to the conductivities of components being separated

## ■ Application of electrophoresis for protein purification

- The last choice for protein purification



# Electrophoresis Principles

## ■ Protein mobility in an electric field

- $Ez = 6\pi\eta rv$  in a steady state
  - $E$  : electric field (volts/m)
  - $z$ : net number of charge
  - $\eta$  : viscosity
  - $r$ : particle radius (Stokes radius)
  - $v$ : velocity
- Specific mobility  $u = v/E = z/6\pi\eta r$

## ■ Free boundary electrophoresis

- Difficult to separate molecules
  - Diffusion
  - Overlapping of components
  - Protein-protein interactions
    - High salt to minimize interaction → heat generation
- Electrophoretic buffer
  - Tris-borate at pH 8-9
    - Low conductivity
    - Most proteins move to anode



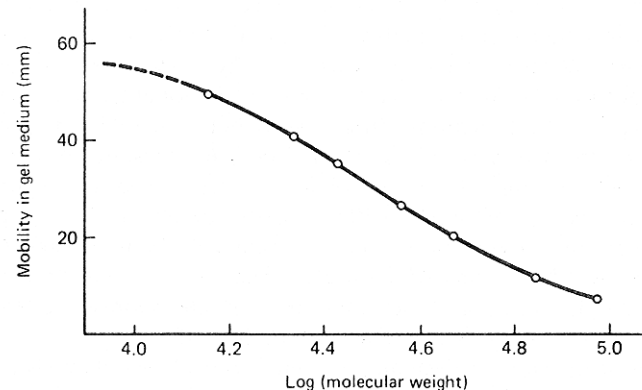
# Electrophoresis Principles

## ■ Gel electrophoresis (polyacrylamide gel)

- Separation by size and charge
  - Good for analysis but technically difficult for preparative work
- Molecular sieving effect
  - Mobility is inversely proportional to logMW for similarly shaped molecules having same charge density

## ■ SDS-PAGE

- $u = u_o(A - \log MW)/A$ 
  - A: logMW of a molecule that would not move in the gel
  - $u_o$ : the mobility of a small molecule unaffected by the gel =  $z/6\pi\eta r$
- $u = z/6\pi\eta r \cdot (A - \log MW)/A$





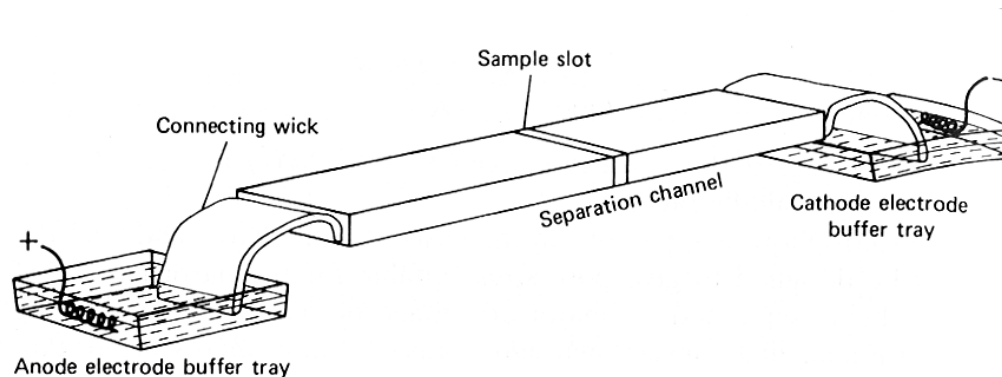
# Preparative Electrophoresis: Horizontal Slabs

## ■ Components of electrophoresis

- Separation channel
- Connection of channel to electrodes via buffer reservoir

## ■ Horizontal slab

- Thick slurry (buffer mixed with starch powder, Sephadex G25 beads etc.) or gel (1~2 % agarose) connected to the buffer reservoir via connecting wick
- Slice the block containing the target protein
- Low resolution but high capacity

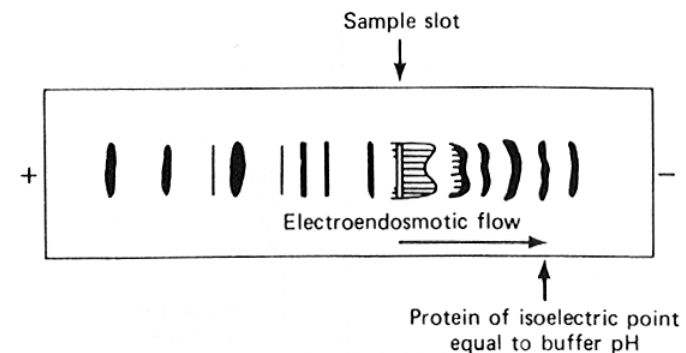
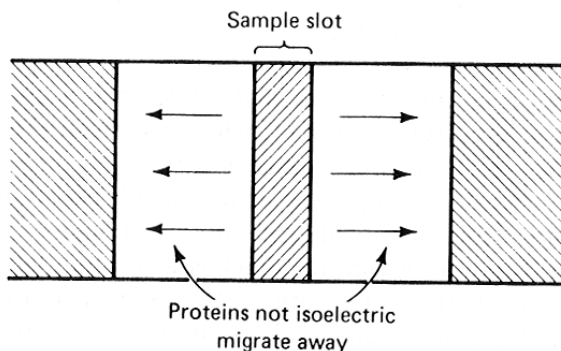




# Preparative Electrophoresis: Horizontal Slabs

## ■ Isoelectric point electrophoresis

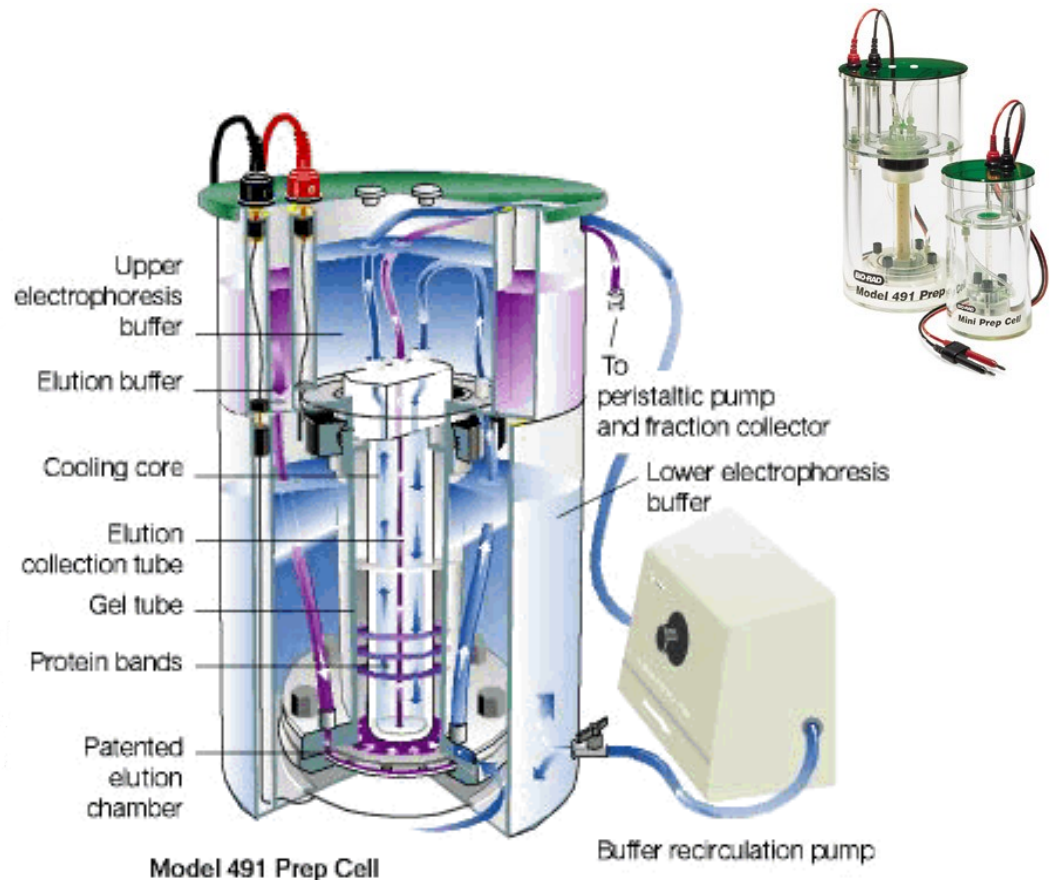
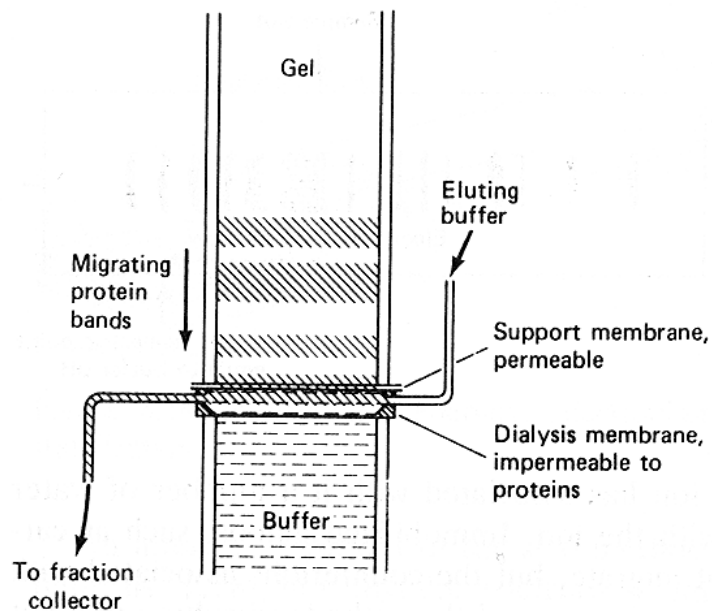
- Run electrophoresis under the conditions of no movement of the target protein
- The pH might be different from isoelectric point determined by isoelectric focusing
  - Depending on the buffer
    - Binding of polyanions → lower the apparent pI
  - Electroendosmotic flow
    - The flow of bulk liquid solvents under the influence of electric field due to immobilized negative charge groups on the walls of electrophoresis channel or gels
    - Movement of associated cation to cathode → movement of water





# Preparative Electrophoresis: Vertical System

- Collection of emerging protein at the bottom of the gel during electrophoresis





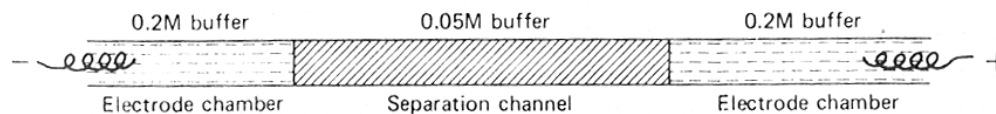
# Buffer Systems for Electrophoresis

## ■ Buffer ions

- Avoid very mobile ions : generate heat
  - Metal ions :  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$
  - Simple anions  $\text{F}^-$ ,  $\text{Cl}^-$ ,  $\text{Br}^-$ ,  $\text{SO}_4^{2-}$ ,  $\text{HPO}_4^{2-}$
- Bulky organic ions with  $n = +1$  or  $-1$ 
  - Slower mobility
- Simple cations
  - Effective in preventing protein-protein interaction

## ■ Buffer systems

- Continuous
  - Same buffer
    - 0.05-0.15 ionic strength, Tris-borate (pH8~8.5) + EDTA
  - Higher concentration in the electrode chambers to lessen voltage losses across the connections



(a) Continuous

- Discontinuous



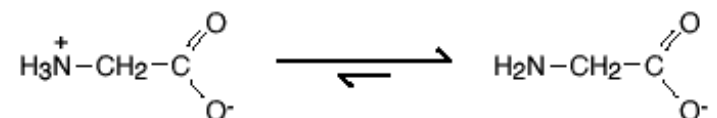
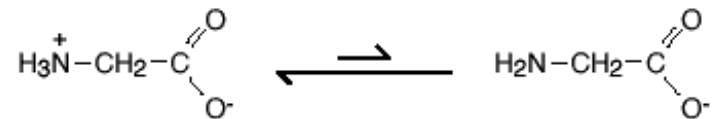
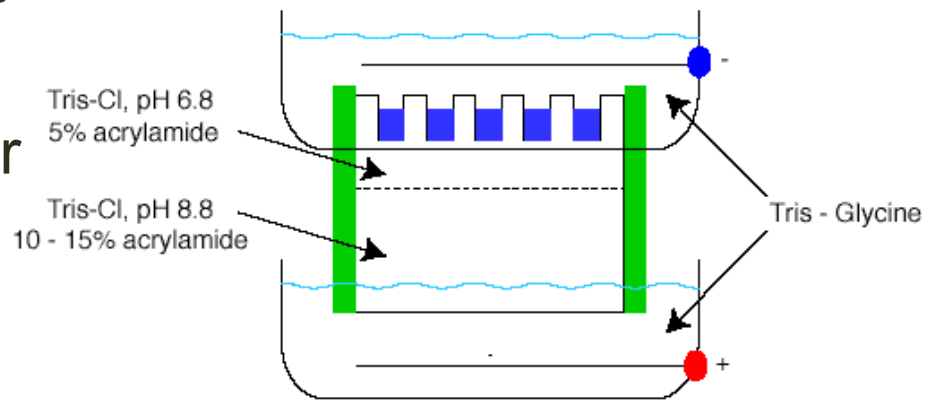
# Discontinuous Buffer Systems

## ■ Principle

- Kohlrausch discontinuity between the starting buffer ions and the following ions with lower mobility
- Band sharpening effect

## ■ Example

- Running buffer
  - Tris-glycine
- Stacking gel
  - Tris-Cl, pH 6.8
  - Low motility of glycine
- Resolving gel
  - Tris-Cl, pH 8.8
  - High motility of glycine

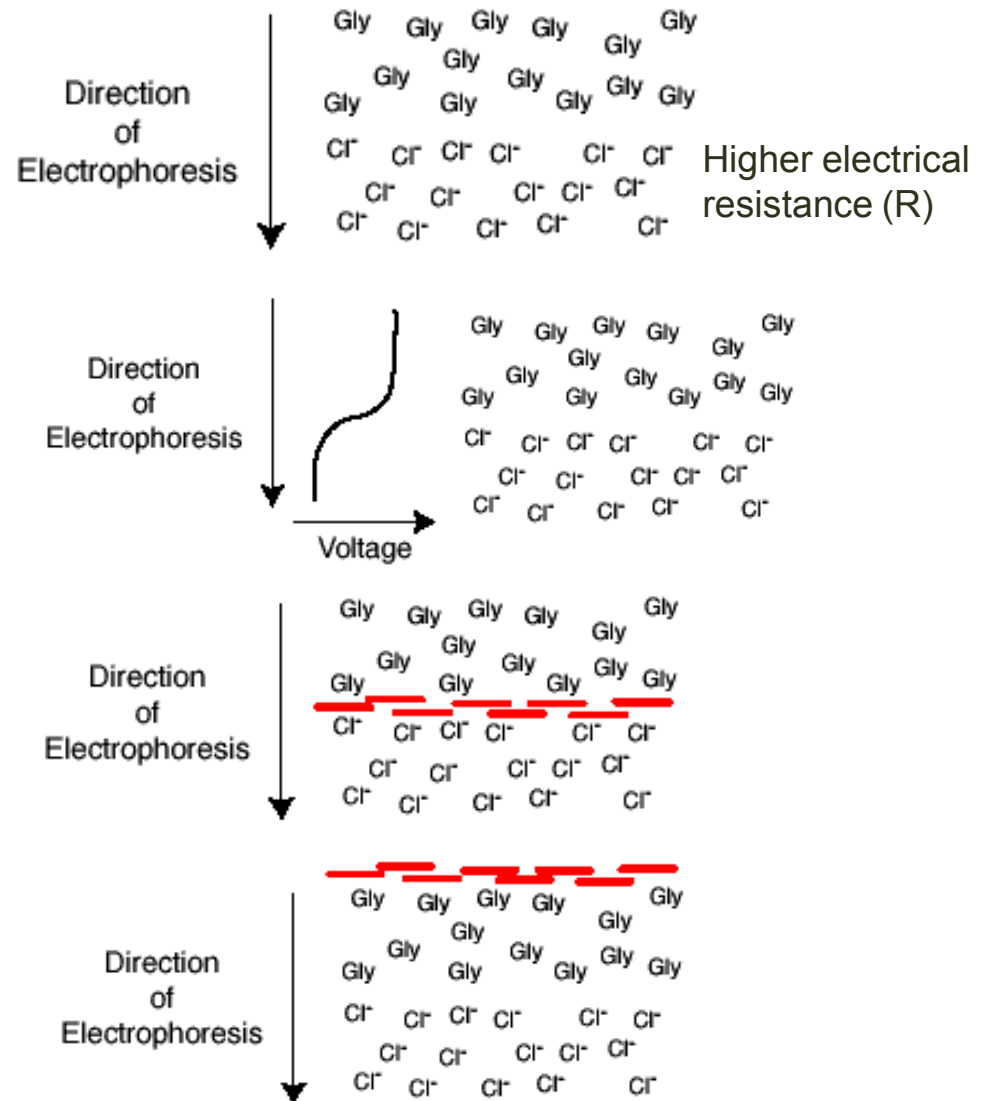




# Discontinuous Buffer Systems

## ■ In stacking gel

- Formation of boundary between glycine and  $\text{Cl}^-$  with higher mobility
- The voltage gradient sharpens the ion boundary
- Proteins are trapped between Gly and  $\text{Cl}^-$



## ■ In resolving gel

- Gly moves faster than proteins
- Separation of proteins depending on charge/mass ratio, size, and shape



# Isoelectric Focusing

## ■ Procedures

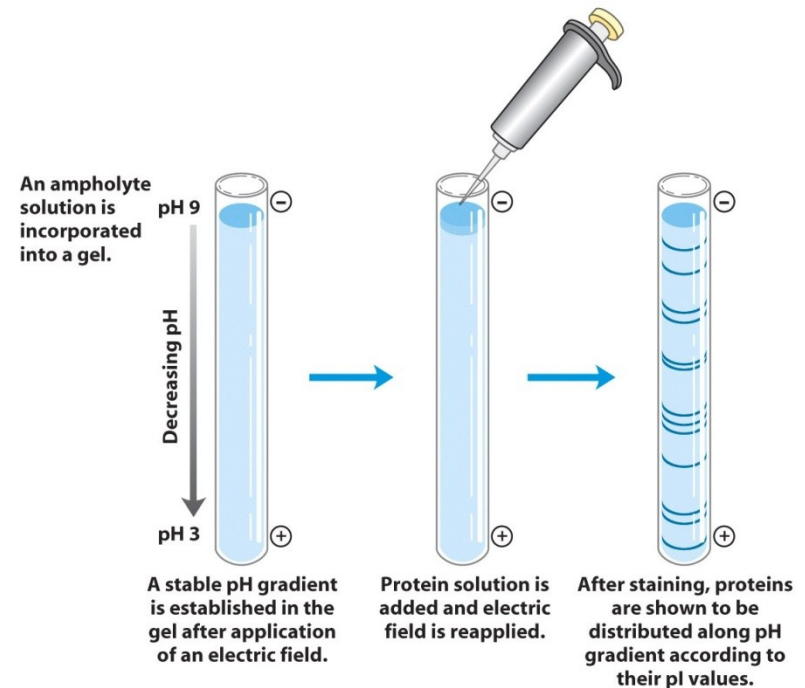
- Establishment of pH gradient
  - Gel containing a mixture of low molecular weight organic acids and bases (ampholytes)
  - Application of electric field
- Each protein migrates until it reaches the pH corresponding to its pI

## ■ Advantage

- High resolution
- High electric field can be applied with little heating

## ■ Requirement

- The protein must be stable at its pI
- The protein must be soluble around its pI even at zero ionic strength







## 3. Liquid Phase Partitioning





# Separation of Proteins by Liquid Phase Partitioning

## ■ Requirement

- Creation of two immiscible liquid phases
- Each liquid phase can dissolve proteins
- Proteins should remain bioactive after dissolving

## ■ Phase separation

- Addition of certain pairs of hydrophilic polymers to aqueous solutions
  - Generation of several phases
- Dextran and PEG
  - Phase separation
    - Dextran-rich (bottom) : Partition of most proteins
    - PEG-rich (top)
  - Useful for large scale production
    - Whole cell lysate can be applied without removing cell debris
    - High capacity : 100mg/ml



# Affinity Partitioning

## ■ Dextran and PEG + PEG-ligand

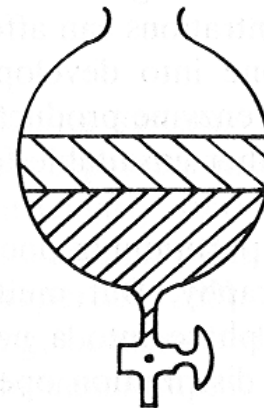
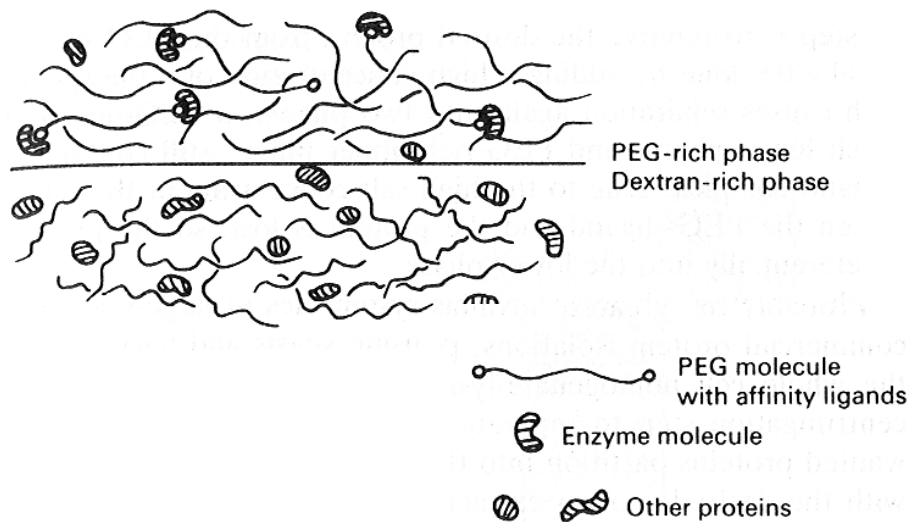
### ■ PEG-ligand

- Activation and attachment of ligand to  $-OH$  of PEG
- Mix with the bulk PEG

### ■ Partitioning of the target protein in the upper phase

### ■ Elution of the protein from the PEG phase

- Addition of high concentration of phosphate buffer
- Phase separation
- Protein partition into phosphate-rich lower phase





# Liquid-Liquid Chromatography

## ■ Immobilization of one liquid phase

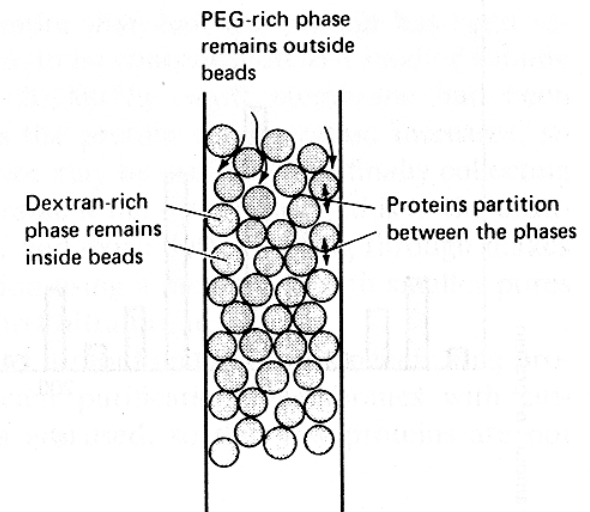
- Equilibrate agarose bead with one of the phases
  - E.g dextran-rich phase
- Use the other phase as moving buffer in column
- Surface tension prevents mixing of two phases

## ■ Problems of phase partitioning methods

- Handling problems with viscous solutions
- Cost : dextran is expensive
  - Alternative: starch derivatives
  - Temperature-dependent phase separation
    - Triton X-114
    - Copolymers of ethylene an propylene oxide

## ■ Advantage

- Gentler than adsorption chromatography
  - Suitable for labile enzymes
- Rapid
- High capacity
- Requires little equipment







## 4. Ultrafiltration





# Ultrafiltration

## ■ Principle

- Forcing liquid through semi-permeable membrane using hydrostatic pressure
  - Molecules smaller than 'cutoff' pore size of membrane → pass
  - Bigger molecules → retain
- Less discriminating than gel filtration

## ■ Applications

- Concentration of protein
  - Cutoff 20 of 10 kDa
- Also can have some purification effect depending on the choice of membrane cutoff size and the molecular weight of the target protein

