



6. Sedimentation

Sedimentation

■ Definition

- The movement of particles or macromolecules in an inertial field

■ Accelerations range

- $1 \times g$ to $100,000 \times g$

■ Applications in bioprocessing

- Clarification of broths and lysates
- Collection of cells and inclusion bodies
- Separation of fluids having different densities



6.1. Sedimentation Principles



Equation of Motion

■ Spherical Particle

- Radius a , density ρ , mass $(4/3) \pi a^3 \rho$

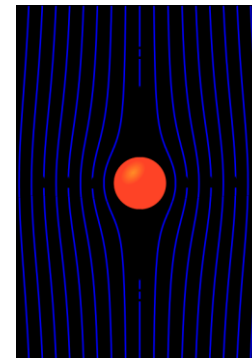
■ Centrifugal acceleration

- $\omega^2 R$
 - ω : angular velocity (rad/s)
 - R : The distance of the particle from the center of rotation

■ Forces in the R direction

- $$m \, dv/dt = F_{iR} + F_{bR} + F_{dR}$$
$$= (4/3) \pi a^3 \rho \omega^2 R - (4/3) \pi a^3 \rho_0 \omega^2 R - 6 \pi \mu a v$$

- i: inertial acceleration
- b: buoyancy due to the density of the medium ρ_0
- d: the Stokes drag force
 - Proportional to the velocity (v) and viscosity (μ) under conditions of creeping flow



Equation of Motion

- Centrifugal velocity
 - At steady state $dv/dt=0$
 - $v = [2a^2(\rho - \rho_0) \omega^2 R] / 9\mu$
- Sedimentation only in the presence of gravity
 - $v = [2a^2(\rho - \rho_0) g] / 9\mu$
 $g = 9.8 \text{ m/s}^2$
- Moving particle
 - R is not constant $v = dR/dt$
 - $dR/R = \{ [2 a^2(\rho - \rho_0) \omega^2 R] / 9\mu \} dt$
 - Integration, at $t = 0, R = R_0$
 - $\ln(R/R_0) = [2a^2(\rho - \rho_0) \omega^2 t] / 9\mu$
 - Relationship between time and travel distance of the particle

Sensitivities

■ Reynolds number

- $Re = 2av\rho/\mu$, typically <0.001

- $\rho_0 = 1.0 \text{ g/cm}^3$, $\mu = 1.0 \text{ cp}$

■ Sedimentation velocities

| Particle | $a \text{ (}\mu\text{m)}$ | $\rho \text{ (g/cm}^3\text{)}$ | Dimensionless acceleration (G), $\omega^2 R/g$ | $V \text{ (cm/h)}$ | Re |
|----------------|---------------------------|--------------------------------|--|--------------------|--------------------|
| Yeast cell | 2.5 | 1.1 | 1 | 0.5 | 7×10^{-6} |
| Bacterial cell | 0.5 | 1.1 | 1 | 0.02 | 6×10^{-8} |
| Protein | 0.05 | 1.3 | 10^4 | 0.06 | 2×10^{-9} |

Sensitivities

- **Isopycnic or equilibrium sedimentation**
 - When $\rho = \rho_0$
 - Used for determination of molecular densities and separation of living cells using density gradient or density shelf
 - E.g. separation of lymphocyte
 - Density shelf with a density around 1.07 g/cm (combination of Ficoll, hypaque)
 - Sedimentation of RBC and floating of WBC

TABLE 5.2

Measured Values of the Density of Representative Cells, Organelles, and Biomolecules

| Cell, organelle, or biomolecule | Density, ρ (g/cm ³) |
|-----------------------------------|--------------------------------------|
| <i>Escherichia coli</i> | 1.09 ^a |
| <i>Bacillus subtilis</i> | 1.12 |
| <i>Arthrobacter</i> sp. | 1.17 |
| <i>Saccharomyces pombe</i> | 1.09 |
| <i>Saccharomyces cerevisiae</i> | 1.11 ^a |
| <i>Amoeba proteus</i> | 1.02 |
| Murine B cells | 1.06 ^a |
| Chinese hamster ovary (CHO) cells | 1.06 |
| Peroxisomes | 1.26 ^a |
| Mitochondria | 1.20 ^a |
| Plasma membranes | 1.15 ^a |
| Proteins | 1.30 ^a |
| Ribosomes | 1.57 ^a |
| DNA | 1.68 ^a |
| RNA | 2.00 ^a |

^aAverage value.

Sensitivities

■ Hindered settling

- Increase in the concentration of sedimenting particles → decrease in v

- $v_c = v(1-\Phi)^n$

- v_c : sedimentation velocity of particles in a concentrated suspension
- v : velocity of individual particles
- Φ : the volume fraction of the particles
- n : a function of the shape of the particle and of the Re
 - » Re < 0.2, $n=4.65$

TABLE 5.3
Effect of Particle Volume Fraction ϕ on the Particle Sedimentation Velocity for Spherical Particles

| ϕ | v_c/v |
|--------|---------|
| 0.01 | 0.95 |
| 0.05 | 0.79 |
| 0.10 | 0.61 |
| 0.20 | 0.35 |



6.2. Methods and Coefficient



Equilibrium Sedimentation

■ Methods to generate concentration gradient

- Step by step layer of solutions of decreasing density
- Centrifugation at extremely high speed
 - Isothermal stratification of a density forming solute like CsCl
- Gradient mixing
 - Linkage of two cylindrical containers with high and low density solutions
 - Time-dependent solute concentration
$$C(t) = c_{1,0} + [Q(c_2 - c_{1,0}) / (2V_0)] t$$
 - » Q : outflow rate, pumping or gravity feed
 - » V_0 : initial volume
 - » c_2 : concentration in the nonmixed chamber (constant)
 - » $c_{1,0}$: Initial concentration in the mixed chamber

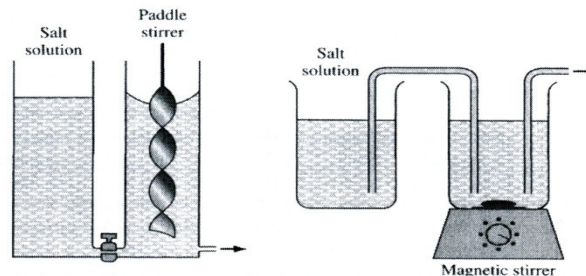


Figure 5.1 Two types of linear gradient mixer.

Sedimentation Coefficient

■ Sedimentation coefficient (s)

- $s = v/(\omega^2 R)$

- $v = [2a^2(\rho - \rho_0) \omega^2 R] / 9\mu$

- $s = 2a^2(\rho - \rho_0) / 9\mu$

- Defined in terms of the properties of the particle and the medium

- $s_{20,w} (s)$

- At 20°C, under conditions of pure water

- $10^{-13}s = 1 \text{ svedberg unit (S)}$

- Svedberg: inventor of the ultracentrifuge

Example : Time required for sedimentation

■ Conditions

- 70S ribosome
- Centrifuge at 10,000 rpm
- $R_o = 4$ cm, $R = 5$ cm

■ Time for complete sedimentation

- $s = v/(\omega^2 R) = dR/dt \cdot 1/(\omega^2 R)$
- Integration, at $t=0$, $R = R_o$
- $\omega^2 s t = \ln (R/R_o)$
- $t = 8.1$ h

Equivalent Time

■ Equivalent time

- Used to assess the approximate properties of a particle type to be separated
- Dimensionless acceleration $G = \omega^2 R / g$
- Equivalent time = $Gt = \omega^2 R / g \cdot t$
 - t : time required for sedimentation
 - Eukaryotic cells: 0.3×10^6 s
 - Bacteria: 9×10^6 s
 - Ribosome : 1100×10^6 s
- Scale up
 - Assume constant equivalent time
 - $(Gt)_1 = (Gt)_2$

Sigma Analysis

■ Sigma Analysis

- Used for engineering analyses and scaling up

■ Q: volumetric flow rate of feed flow

- $Q = \{v_g\} [\Sigma]$

- $v_g = 2a^2(\rho - \rho_0)g / 9\mu \quad (1)$

- Sedimentation velocity at 1 x g
- Properties of the particle and the fluid

- Σ

- Represents the geometry and speed of the centrifuge
- Cross-sectional area equivalent of the centrifuge
- Properties of the centrifuge

- $\ln(R/R_0) = [2a^2(\rho - \rho_0) \omega^2 t] / 9\mu \quad (2)$

- $v_g = [g \ln(R/R_0)] / \omega^2 t \quad (1) + (2)$

- Can be determined in the lab
 - t: the minimum time to clarify the sample
 - R: distance from the center of rotation to the top of the packed solids
 - R_0 : distance from the center of rotation to the top of the liquid



3. Production Centrifuges



Common Types of Production Centrifuge

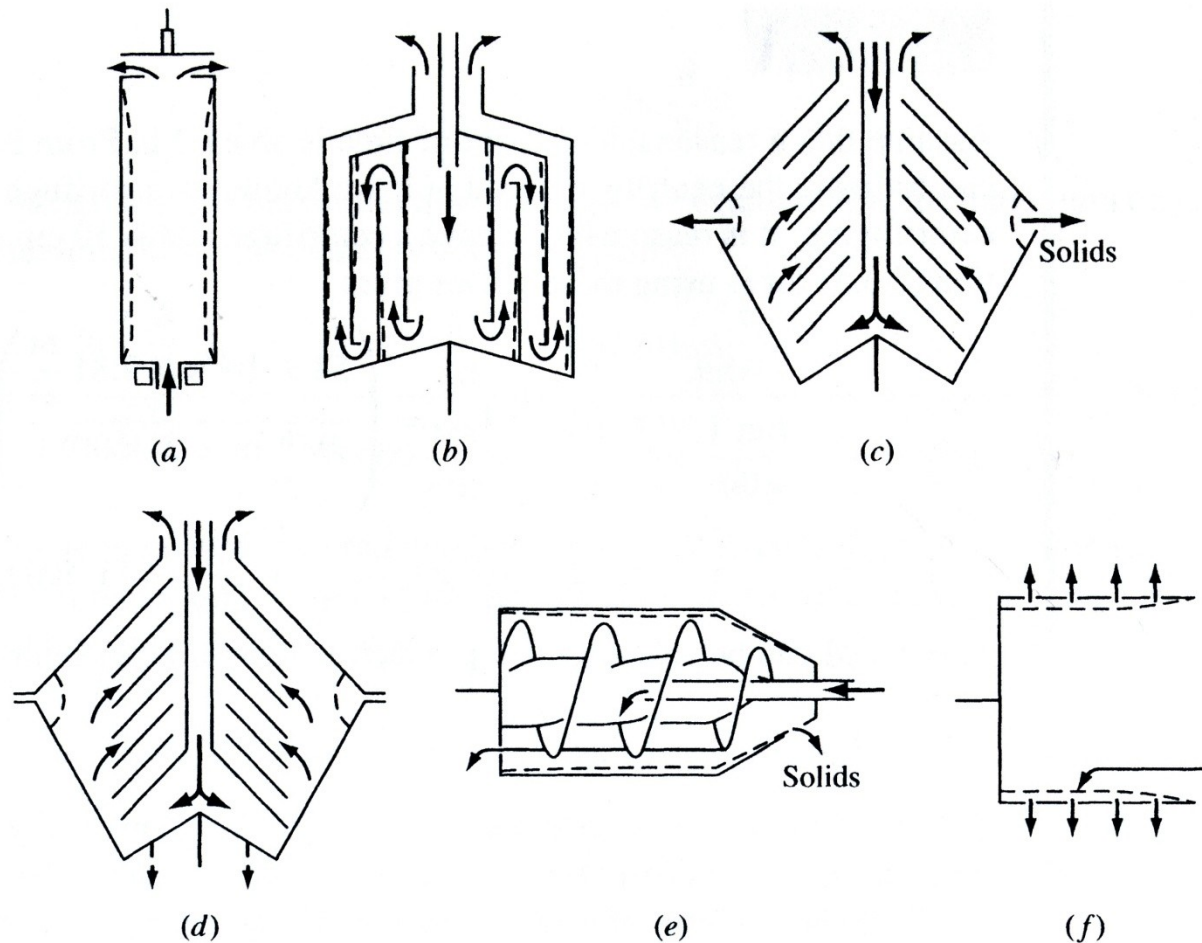


Figure 5.2 Common types of production centrifuge: (a) tubular bowl, (b) multichamber, (c) disk, nozzle, (d) disk, intermittent discharge, (e) scroll, and (f) basket. Arrows indicate the path of the liquid phase; dashed lines show where the solids accumulate.

Common Types of Production Centrifuge

■ Tubular centrifuge

- For particles of relatively low sedimentation coefficient
- e.g. protein precipitate

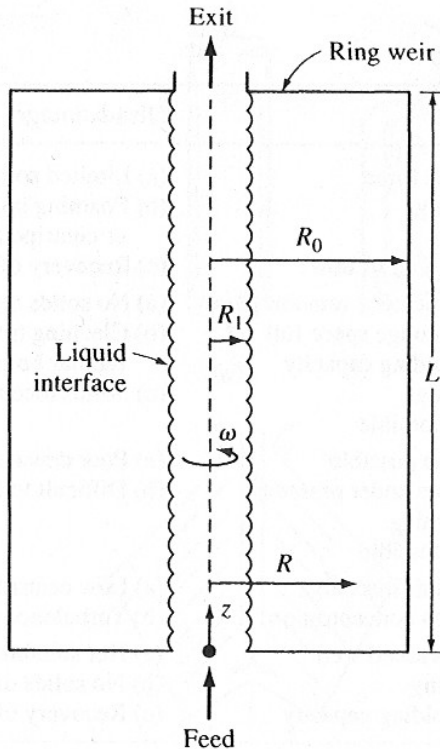
■ Disk centrifuge

- Large sedimentation area
- Continuous or intermittent solids discharge is possible
- e.g. Cells and cell lysates

■ Scroll and basket centrifuges

- For particles of relatively high sedimentation coefficient
- e.g. Antibiotic crystal

Tubular Bowl Centrifuge



Q: flow rate

- Trajectory of sedimented particles
 - In the radial direction (1)
 - $dR/dt = [2a^2(\rho - \rho_0) \omega^2 R] / 9\mu$
 - In the axial direction (2)
 - $dz/dt = Q/A = Q / [\pi (R_0^2 - R_I^2)]$
- Combination of (1) and (2)
 - $(dR/dt) / (dz/dt) = dR/dz$
 - Integration
 - $Q = \{ [2a^2(\rho - \rho_0)] / 9\mu \} [\pi L (R_0^2 - R_I^2) \omega^2 / \ln (R_0 / R_I)]$
 - $Q = \{ v_g \} [\Sigma]$
 - $v_g = 2a^2(\rho - \rho_0)g / 9\mu$
 - $\Sigma = \pi L (R_0^2 - R_I^2) \omega^2 / [g \ln (R_0 / R_I)]$
- Practical usage
 - To get an optimal centrifuge
 - Determine v_g using a benchtop centrifuge
 - Calculate Q from process requirement
 - Determine Σ to get a proper centrifuge



4. Ultracentrifugation



Ultracentrifugation

■ Ultracentrifugation

- 50,000 to 100,000 g
- Very small particles and macromolecules can be sedimented

■ Types

- Analytical centrifugation
 - < 0.1 ml volume sample
 - Monitoring of centrifugation in an optical cell
- Preparative centrifugation
 - Up to 50 ml sample
 - Collection of the sample by puncturing the bottom of the centrifuge tube

Determination of Molecular Weight

- Determination of molecular weight by the combined measurement of sedimentation and diffusion coefficient
- $m \, dv/dt = F_{iR} + F_{bR} + F_{dR}$
- $V(\rho - \rho_0) \omega^2 R - 6\pi\mu a v = 0$ at steady state
 - V : volume of a single molecule
- If $V = m\bar{V} = m/\rho$
 - $m = 6\pi\mu a v / [(1 - \bar{V}\rho_0) \omega^2 R]$ (1)
 - m : mass of a single molecule
 - \bar{V} : specific volume
 - Stokes-Einstein equation:
 - $\mathcal{D}\mu/kT = 1/6\pi a$
 - $6\pi a \mu = kT / \mathcal{D} = RT/\mathcal{N}\mathcal{D}$ (2)
 - k : Boltzmann's constant = R (gas constant)/ \mathcal{N} (Avogadro number)
 - $s = v/(\omega^2 R)$ (3)
- $M = sRT/\mathcal{D}(1 - \bar{V}\rho_0)$: Substituting (2) and (3) into (1)
 - M : molecular weight